

# ***In vitro* androgenesis in pepper (*Capsicum annuum* L.) and the affecting factors on success: II. Carbohydrate source and antioxidants**

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## **Abstract**

Microspore cells develop into male gametophytes, which are released as pollen. Under certain stress conditions, the developmental pathway of microspores can be transformed into an embryo instead of pollen with haploid technology. In this experiment, 2 pepper breeding lines (G-1 and G-2) and 4 nutrient media formed with Murashige & Skoog (1962-MS) + 30 g L<sup>-1</sup> sucrose or maltose and with or without vitamins as antioxidants. 0.05 mg L<sup>-1</sup> biotin (vitamin B7) and 0.5 mg L<sup>-1</sup> ascorbic acid (vitamin C) were studied as antioxidants. The anthers were pretreated for 2 days at 35 °C in dark, then they were incubated in a climate chamber at 25 °C and 16/8 hours photoperiodic conditions. The highest embryos rate, and development of embryos into the plantlet has been obtained from the medium containing maltose, and antioxidants. Although there was no significant differences between genotypes in medium- I (MS + sucrose), medium-II (MS + sucrose and antioxidants) and medium-III (MS + maltose) in terms of embryogenic response, a significant difference was determined between genotypes in medium-IV (MS + maltose and antioxidants). The number of embryos obtained from the G-1 in medium-IV has increased 2.5, 6.4, and 4.5 times, compared to the medium-I, medium-II, and medium-III respectively.

## **Introduction**

Doubled haploidy (DH) system is a widely used tool in plant breeding, genetic research, and biotechnology today. The developmental pathway of a gamete cell can be changed and directed towards sporophytic development instead of mature pollen from microspore with double haploid technology. Accordingly, a gamete cell can form a haploid embryo alone, under certain stress conditions ([Segui-Simarro & Nuez, 2008](#); [Shariatpanahi et al., 2006](#); [Perez-Perez et al., 2019](#); [Testillano, 2018, 2019](#); [Cengiz & Korkut, 2020](#)). Stress

pretreatments applied to anthers and microspores to inducing the sporophytic pathway are inhibit the gametophytic pathway and increase embryogenic potential ([Sanchez et al., 2020](#)). Microspores-induced embryo formation is called androgenesis.

In pepper anther culture, 35°C temperature pretreatment of anthers is the most commonly used stress condition. However, the levels of intracellular reactive oxygen species increase in microspores exposed to high temperature stress conditions and negatively affect the viability of microspores ([Zur et al., 2009](#); [Varnier et al., 2009](#); [Gill & Tuteja, 2010](#)).

Antioxidants reduce the oxidative stress in the plant and ensure the viability of microspores. Vitamin C (ascorbic acid) and vitamin B7 (biotin) are excellent free radical scavengers. They can provide tolerance to osmotic and oxidative stress in plants (Roje, 2007).

In *Capsicum annuum* L.; although different protocols have been employed to increase the productivity of anther culture during the 48 years since the first successful androgenic embryogenesis reported by Wang et al. (1973), George & Narayanaswamy (1973), and Kuo et al. (1973), both embryo productivity and plantlet regeneration rate still have not reached the desired level.

Developing a successful protocol for producing DH plants in each plant species and variety is difficult and may take years. It is known that various factors such as donor plant genotype and growth conditions, culture methods, composition and hormonal composition of the nutrient medium, stress pretreatment that induces embryogenesis, and flower bud development period affect embryo productivity. Although protocols for the production of DHs exist for many species, conditions suitable for one species or one genotype may not work for another (Segui-Simarro & Neuz, 2008; Irikova et al., 2011; Cömlekçioğlu & Ellialtıoğlu, 2018). Therefore, these factors need to be studied in detail in order to precisely determine the combination that works for the donor plant genotype studied.

In order to encourage microspores to embryogenic development even in recalcitrant species, new targets need to be determined and new strategies should be devised. Therefore, in this study, the effects of two factors on the yield of haploid embryo, the carbon source (sucrose and maltose) used in the nutrient medium and the use of antioxidants (ascorbic acid and biotin), were investigated in two *copia* type pepper breeding lines.

## Materials and Methods

This study was carried out in the greenhouse and tissue culture laboratories of United Genetics Vegetable Seeds Company (Mustafakemalpaşa, Bursa, Turkey). Two *copia* type pepper (*Capsicum annuum* L.) breeding lines of the company coded as G-1 and G-2 were used as the donor plant. Donor plants were grown in an unheated greenhouse. Donor plant seedlings were planted in the unheated greenhouse of United Genetics Turkey Vegetable Seeds Company in Mustafakemalpaşa/Bursa location on April 13, 2020. Flower bud harvesting for anther culture started on 20 May and continued until June 15. Table 1 shows the certain climatic variables of greenhouse for the experimental year for the months which experiments were conducted (from donor plants planting to the end of flower buds harvested). According to their morphological development, the buds in the developmental stage where the corolla, and calyx are the same height or the corolla height is slightly above

the calyx height were collected and brought to the laboratory. Anthers at this developmental stage mostly have late uninucleate, and middle uninucleate microspores or are known to contain microspores in the young binucleate stage (Cömlekçioğlu & Ellialtıoğlu, 2018).

**Table 1.** Temperature and relative humidity conditions of greenhouse during the experiments

Year (2020)	Minimum temperature (°C)	Average air temperature (°C)	Max. air temperature (°C)	Average relative humidity
April	13	24.0	35	48.6
May	16	26.5	37	47.9
June	17	27.0	38	54.2

For surface sterilization, flower buds were first rinsed with water, then with 70% ethanol for 10-15 seconds and kept in 10% commercial bleach (containing 5% sodium hypochlorite) and a drop of Tween-20 for 10 minutes. It was washed 3 times with sterile distilled water. The buds were carefully cut and isolated in a laminar flow cabinet on sterile paper.

In the experiment, four MS (Murashige & Skoog, 1962) culture media, supplemented with sucrose or maltose, and with or without 5 mg L<sup>-1</sup> ascorbic acid, 0.05 mg L<sup>-1</sup> biotin combination (as antioxidants) were used. In our previous study, 15 g L<sup>-1</sup>, 30 g L<sup>-1</sup>, and 60 g L<sup>-1</sup> maltose were tested and the best result was obtained from 30 g L<sup>-1</sup>. Therefore, 30 g L<sup>-1</sup> maltose was studied in this study (Bat et al., 2020). All chemicals used in this experiment were Duchefa Biochemie brand. The culture media used are:

Medium-I (M-I); MS + 4 mg L<sup>-1</sup> naphthalene acetic acid (NAA), 0.5 mg L<sup>-1</sup> benzyl amino purine (BAP), 0.25% activated charcoal (AC) 15 mg L<sup>-1</sup> silver nitrate (AgNO<sub>3</sub>), 7 g L<sup>-1</sup> agar, 30 g L<sup>-1</sup> sucrose,

Medium-II (M-II); MS + 4 mg L<sup>-1</sup> NAA, 0.5 mg L<sup>-1</sup> BAP, 0.25% AC, 15 mg L<sup>-1</sup> AgNO<sub>3</sub>, 30 g L<sup>-1</sup> sucrose, 7 g L<sup>-1</sup> agar, 0.5 mg L<sup>-1</sup> ascorbic acid, 0.05 mg L<sup>-1</sup> biotin,

Medium-III (M-I); MS + 4 mg L<sup>-1</sup> NAA, 0.5 mg L<sup>-1</sup> BAP, 0.25% AC, 15 mg L<sup>-1</sup> AgNO<sub>3</sub>, 7 g L<sup>-1</sup> agar, 30 g L<sup>-1</sup> maltose,

Medium-IV (M-IV); MS + 4 mg L<sup>-1</sup> NAA, 0.5 mg L<sup>-1</sup> BAP, 0.25% AC, 15 mg L<sup>-1</sup> AgNO<sub>3</sub>, 7 g L<sup>-1</sup> agar, 30 g L<sup>-1</sup> maltose, 0.5 mg L<sup>-1</sup> ascorbic acid, 0.05 mg L<sup>-1</sup> biotin.

Anthers planted on nutrient media were exposed to heat pre-treatment at 35°C under continuous darkness for the first 2 days. It was then incubated at 25°C under photoperiod conditions for 16/8 hours.

## Statistical Analysis

The experiment was carried out in a randomized block design with 3 replications. At each replication, 100 anthers were cultured in 10 Petri dishes containing 10 anthers each. The data were subjected to analysis of variance (ANOVA) using Tarist (Açıkgöz et al., 2004). Mean separation was performed by Fisher's Least Significance Difference (LSD) (p<0.01).

**Table 2.** Summary of ANOVA for total embryos obtained, embryo rate and regenerated plantlets

Source of variation	DF	Total embryos		Embryo rate (embryo per 100 anther)		Number of regenerated plantlets	
		Mean square	F-value	Mean square	F-value	Mean square	F-value
Genotype (G)	1	70.04	175.63**	81.29	154.67**	4.10	21.18**
Media (M)	3	160.82	403.25**	207.04	393.93**	30.23	156.31**
GxM	3	43.60	109.32**	30.01	57.09**	19.63	101.52**
Error	14	0.39		0.53		0.19	
Total	23	30.03		34.86		7.45	

\*Significant at alpha level 5%; \*\* Significant at alpha level 1%; DF= Degrees of freedom

## Results and Discussion

This study was conducted to examine the androgenesis response of pepper to the carbohydrate source (sucrose or maltose) and 'biotin + ascorbic acid' supplementation as antioxidants in the nutrient medium. In this study, 4 different nutrient media were tested. Variance analysis results are presented in Table 2. Approximately 2 months (58 days) after the anthers were cultured, the first embryos were seen and embryo formation continued for almost 5 months.

The number of embryos obtained showed significant differences according to genotype, nutrient medium, and 'genotype × medium' interaction. Both genotypes produced a higher number of embryos in M-I and M-IV compared to the M-II, and M-III. While G-I produced the highest number of embryos in M-IV, gave a similar number of embryos in M-II and M-III. G-2 produced a higher number of embryos in M-I and M-IV compared to the M-II, and M-III. In the M-IV, the difference between the 2 genotypes was found to be very significant (Table 3). In this experiment, it was determined that the genotype effect was more important than the nutrient medium.

Genotype × medium interaction was found important in terms of androgenesis rate (embryo obtained per 100 anthers). The highest rate of the embryo was obtained from G-1 in M-IV. Both genotypes had the lowest values in M-II containing sucrose, and ascorbic acid, and biotin. It has been determined that adjusting the most suitable nutrient medium according to the genotype is important in androgenesis.

Figure 1 shows a few microspore-derived embryos and fully developed green plantlets obtained from this experiment. While some of the embryos obtained from the culture develop into a complete plantlet, some embryos cannot form green plants. Accordingly, it was

determined that the number of plantlets developed from the obtained embryos differed significantly in terms of genotypes, media, and their interactions. The lowest plantlet growth was determined in the M-II and the highest in M-IV, parallel to the number of the embryos obtained (Table 3).

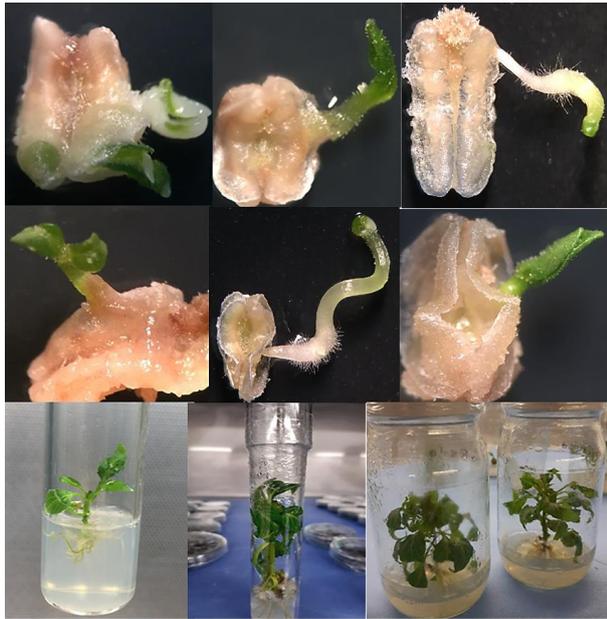
The results showed that the interaction of ascorbic acid, and biotin combination with different carbohydrate sources is important. While determining the negative effects of ascorbic acid and biotin combination on embryogenesis in sucrose contained medium, it has been observed that ascorbic acid and biotin combination has a significant positive effect in maltose content medium. The type of sugar used in the nutrient medium had a significant effect on androgenesis, and maltose had a positive effect on inducing haploid embryo formation. The combination of 30 g L<sup>-1</sup> maltose, 0.5 mg L<sup>-1</sup> ascorbic acid, and 0.05 mg L<sup>-1</sup> biotin was the best culture medium for induction of high-frequency embryogenesis (26.1% for G-1 and 16.4% for G-2). Both genotypes gave similar results in terms of embryogenic response in M-I, M-II, and M-III, whereas a significant difference was detected between genotypes in medium-IV. The number of embryos obtained from G-I increased 2.5, 6.4, and 4.5 times, compared to the M-I, M-II, and M-III respectively. The effect of the culture medium on the success of anther culture in interaction with the genotype was clearly demonstrated in the study.

[Cheng et al.](#) (2020) reported that stress pretreatments to anthers are necessary to induce embryo development from microspores, and heat stress applications in peppers are indispensable since plant regeneration cannot be achieved without heat pretreatment. Stress applications used to reprogram microspores for embryogenesis often cause cell death.

**Table 3.** Total number of embryos, embryo ratio and number of plantlets obtained according to nutrient media and genotypes

Medium	Number of embryos			Embryo rate (Embryo no/100 anthers)*			Number of the plantlets		
	G-1	G-2	Mean	G-1	G-2	Mean	G-1	G-2	Mean
M-I	7.7 b	8.3 a	8.0 b	16.1 b	16.8 a	16.4 b	3.0 b	4.3 a	3.7 b
M-II	3.0 c	1.0 c	2.0 d	9.9 d	5.7 d	7.9 d	1.0 c	1.0 c	1.0 c
M-III	4.3 c	3.3 b	3.8 c	12.0 c	10.5 c	11.2 c	1.3 c	1.7 c	1.5 c
M-IV	19.3 a	8.0 a	13.7 a	26.1 a	16.4 a	21.3 a	9.3 a	3.0 b	6.2 a
Mean	8.5 a	5.2 b		16.0 a	12.4 b		3.7 a	2.8 b	
LSD (%)	G: 0.8, M: 1.1, G × M: 1.6			G: 0.9, M: 1.3, G × M: 1.8			G: 0.6, M: 0.8, G × M: 1.1		

G: Genotype, M: medium; GxM: GxM Interaction. Means followed by the same letter in column do not differ according to least significance difference (LSD) test (P ≤ 0.01). \*Arcsine transformation was performed for analysis of variance



**Figure 1.** Some of microspore-derived embryos and plantlets obtained from the cultures.

It is the main factor that greatly reduces the efficiency of the method in transition to embryogenesis (Testillano, 2018; Rodriguez-Serrano et al., 2012). It limits the use of these methods in many plants because of low totipotens, and embryogenesis, thus appropriate stress conditions must be applied. Stress applied for the induction of embryogenesis causes an increase in reactive oxygen species (ROS) in microspore cells, and death (Zur et al., 2009; Rodriguez-Serrano et al., 2012). In order to prevent oxidative damage in the cell and to control cell death, ROS cleaning applications are of great importance. ROS scavenging roles of vitamins in plant cells have been reported previously (Becker et al., 2014; Zur et al., 2009; El-Sharabasy, 2019).

It has been reported that the vitamins in the culture medium increase the microspore viability and thus the embryogenesis response improves (Al-Khayri, 2001; Roje, 2007; Habibi et al., 2009; Hoseini et al., 2013; Zeng et al., 2015; Ozsan & Onus, 2017). It has been reported that it is important to increase the tolerance of microspore cells against oxidative stress caused by the stress pretreatments applied to anthers. Due to antioxidant properties of certain vitamins in the nutrient medium is a factor that increases the androgenic embryo yield (Demirkaya & Comlekcioglu, 2021). Navarro-Alvarez et al. (2006) reported that carbohydrates are critical components in anther culture medium for successful somatic embryo initiation, and plant regeneration. He reported that galactose and mannose did not support embryo formation, while glucose gave more positive results than sucrose and fructose. However, the highest rate of embryos in three wheat genotypes was obtained from the medium containing maltose.

In pepper anther culture, sucrose is the most commonly used carbohydrate source in culture media

(Taskin et al., 2011; Cömlekçioglu & Ellialtıođlu, 2018). However, there are studies reporting that maltose as a carbohydrate source is more effective for androgenesis compared to sucrose. Last et al. (1990) reported that when maltose is used instead of sucrose in wheat anther culture, a three to four-fold increase in embryo yield was observed although it varied according to genotypes. Trejo-Tapia et al. (2002) found that maltose is the most effective carbohydrate in callus induction. Bat et al. (2020) compared 15 g L<sup>-1</sup>, 30 g L<sup>-1</sup>, and 60 g L<sup>-1</sup> maltose doses with 30 g L<sup>-1</sup> sucrose in a study with 3 pepper genotypes. While emphasizing the importance of the genotype effect in the androgenetic response, the study results also showed that a significantly higher embryo formation was achieved in a medium containing 15 g L<sup>-1</sup> and 30 g L<sup>-1</sup> maltose compared to a medium containing 30 g L<sup>-1</sup> sucrose.

In a small number of trials using maltose (Irikova et al., 2011), maltose was generally used alone and different carbohydrate sources were not compared in the studied genotypes. Gebolođlu et al. (2017) reported that the use of honey instead of sucrose in eggplant anther culture has a positive effect on obtaining embryos, with the highest number of embryo formation being 18.35% and 20.95% from honey and sucrose treatment, respectively.

Although pepper is one of the most intensively studied plants on androgenesis, DH production in this species is still not at the desired level. The relationship between androgenetic response and nutrient medium composition in peppers has been widely accepted. However, although it is known that carbohydrate sources and doses in the nutrient medium affect embryogenesis, studies have focused on growth regulators and the relationship of their doses with the embryogenic potential of microspores rather than determining the most appropriate carbohydrate source, and doses.

It has been determined that the applications are very promising in terms of increasing the yield of haploid embryos with the anther culture method, as well as on the rate of obtaining plantlets from the formed embryos. It is thought that different carbohydrate sources and doses should be studied in also other recalcitrant species to increase the efficiency of haploid embryogenesis, and plantlet regeneration. While emphasizing the importance of the genotype effect once again in the study, it was determined that different factors should be studied in detail in pepper anther culture. Genotype effect in anther culture response is an important factor limiting the application of this technology in plant breeding. This requires the experimental development of culture media, and protocols for different genotypes. Culture protocols that can be successfully applied in different genotypes need to be developed, and optimized.

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## Author Contributions

Conceptualization, writing-review and editing: NC, SSE; Methodology: FNA, MAY, ED; Data curation, Formal analysis: ED.

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